



Defining a gene function by mutational studies in plant

Overview of the lecture

Brief background on molecular biology DNA, genetic materials, mechanism of

transfer of genetic information

How to elucidate the gene in the mutant plant

Making the audience aware about the importance and power of molecular biology in enhancing the quality of our life







Plants are built with the same genetic material that we are made of



Like human or animals, they have chromosomes, where all the genetic information are stored

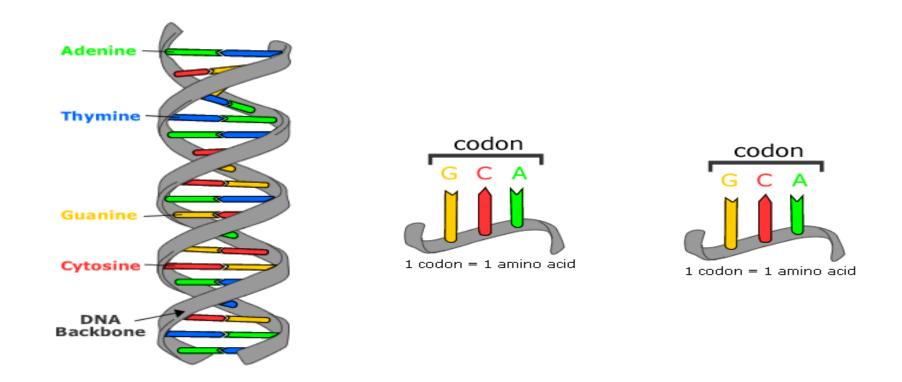




Finally, like human or animal the plant also respond to different stimului such as light, temperature, gravity etc.







Deoxyribonucleic acid, or **DNA**, is a nucleic acid that contains the genetic instructions used in the development and functioning of all known living organisms





More information on DNA

The main role of DNA molecules is the long-term storage of information

DNA contains the instructions needed to construct other components of cells, such as proteins and RNA molecules. It is often called the blue print of life

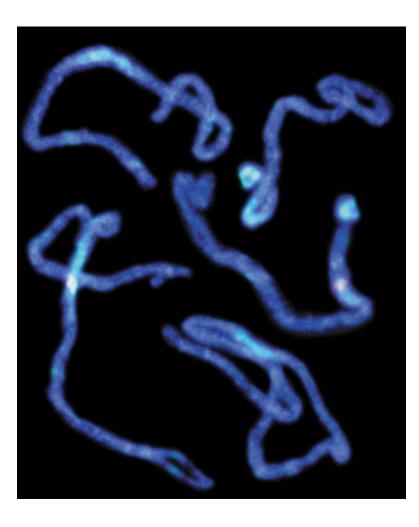
The DNA segments that carry this genetic information are called genes

The other DNA sequences have structural purposes, or are involved in regulating the use of this genetic information





Within cells, DNA is organized into structures called chromosomes and the set of chromosomes within a cell make up a genome

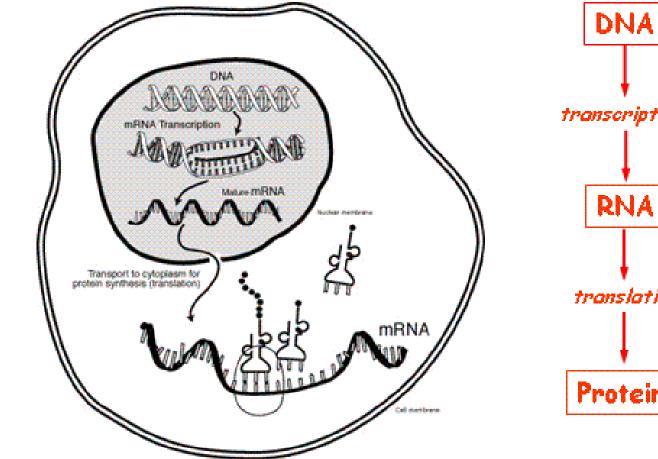


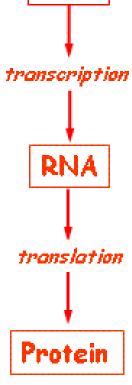
Arabidopsis chromosome





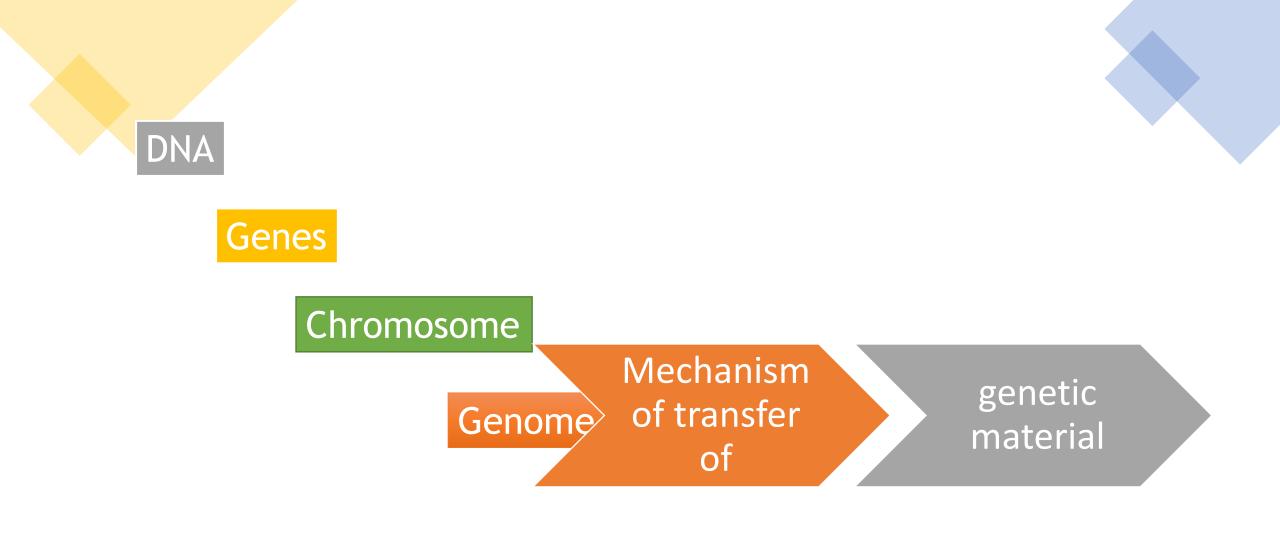
Mechanism of transfer of genetic information















Plants response to different environmental stimului



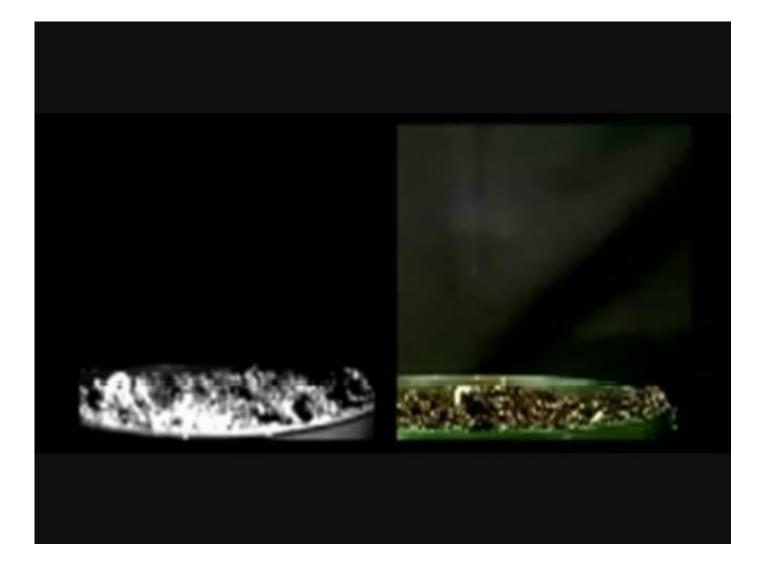




Arabidopsis germination







Germination of Sunflower in light and dark



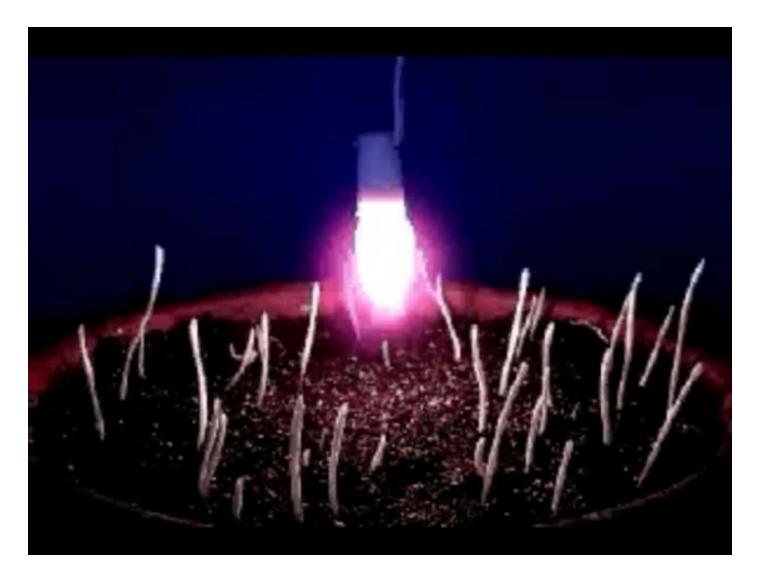




Corn phototropism





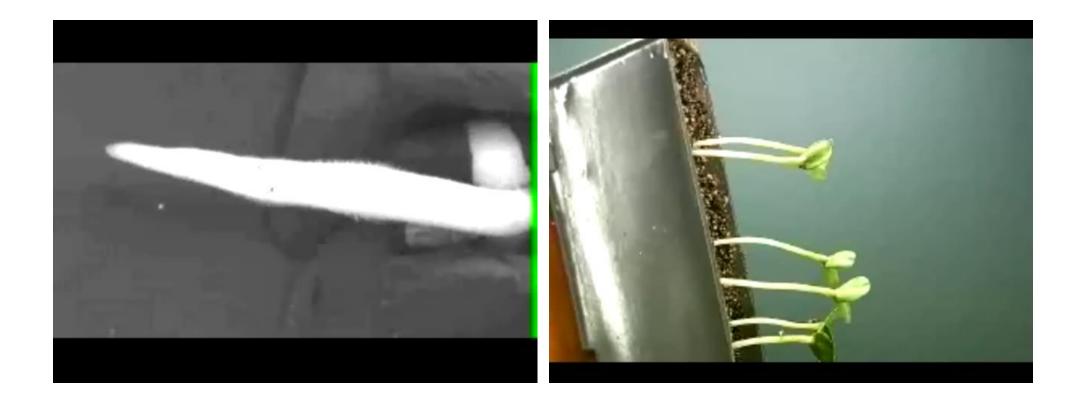


Response to light





Gravity response in root and shoot









Sunflower solar track







1) What determines such responses of plants?

2) Is it possible to find out the factors that influence the plant responses to different stimulus?





Model Plant

- \star Arabidopsis thaliana has a small genome size
- \star The whole genome has been sequenced
- \star The genome of this plant is composed of 5 chromosomes which contains 125 Mb of DNA and 25,948 identified proteins
- It has a shorter life cycle relative to other plants; Takes only six weeks from germination to produce seeds
- It is easy to grow. Virtually it grows everywhere
- Finally transferring the gene in this plant is relatively easy compared to other plants

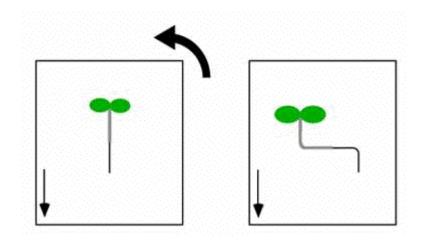
Arabidopsis thaliana

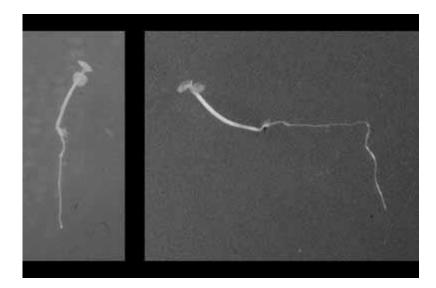
Arabidopsis chromosomes

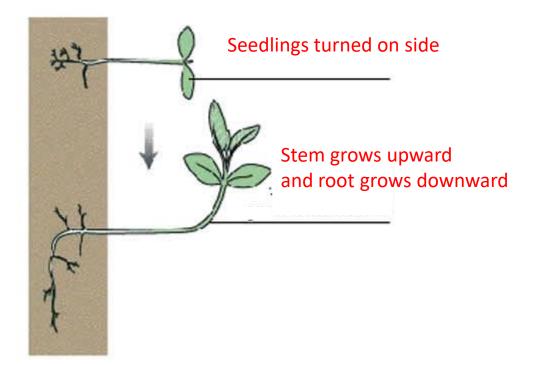






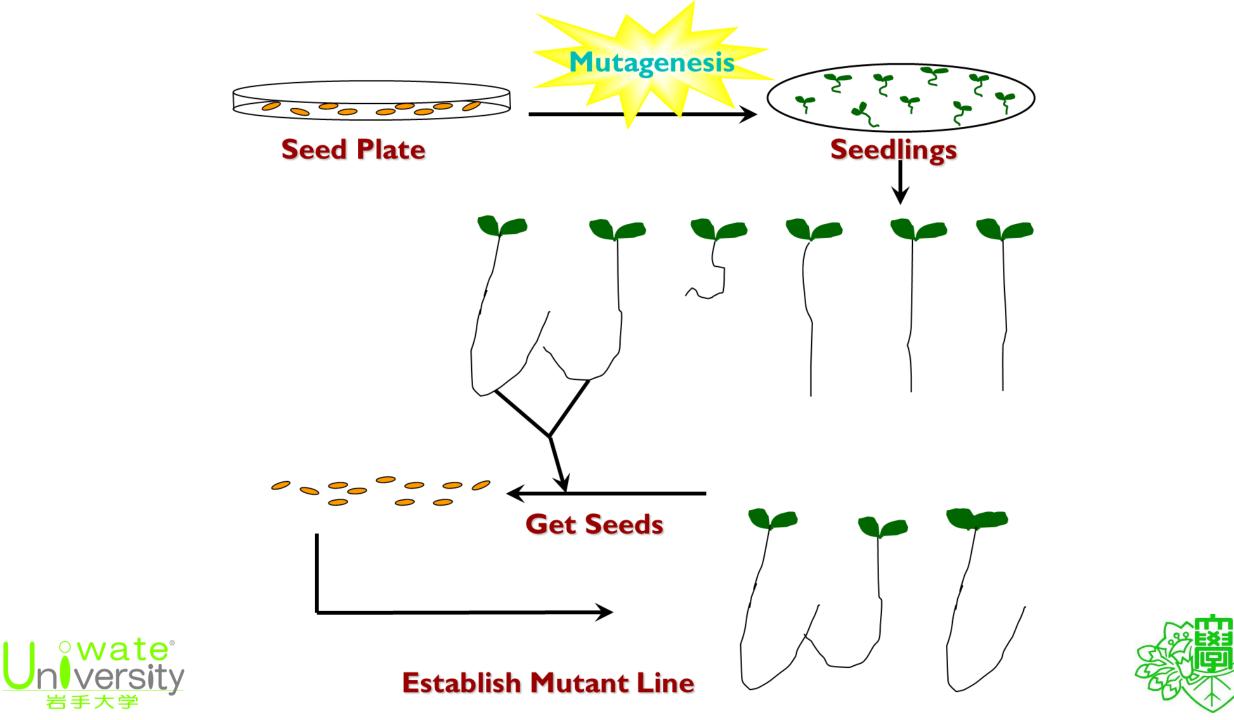










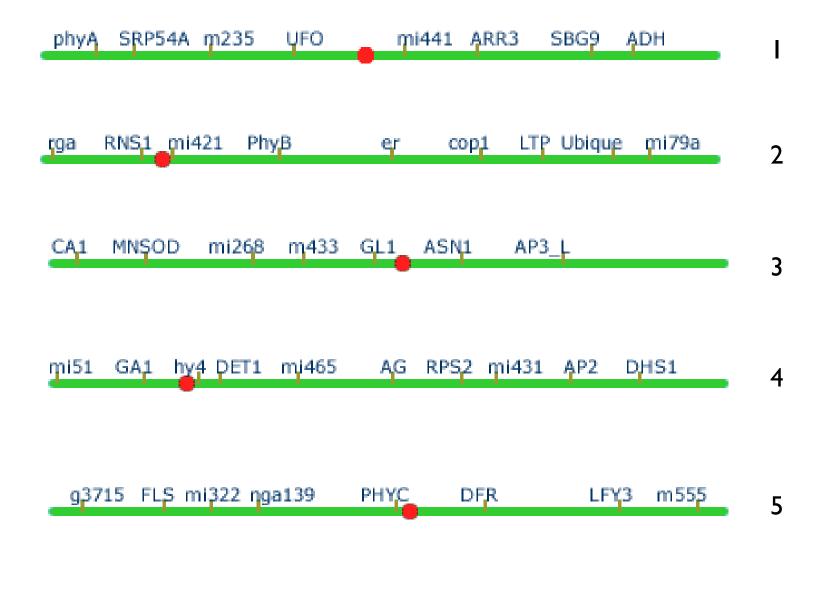








Chromosome



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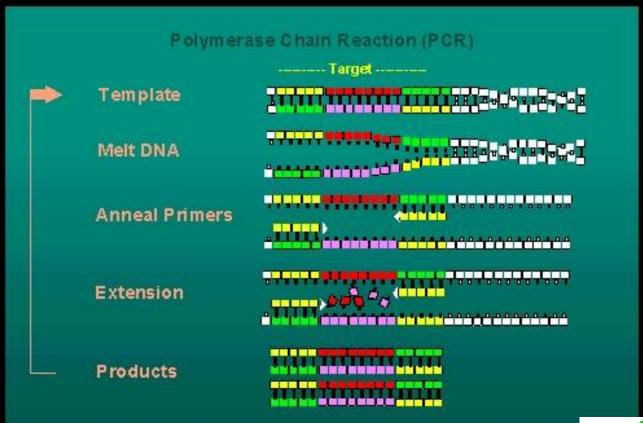
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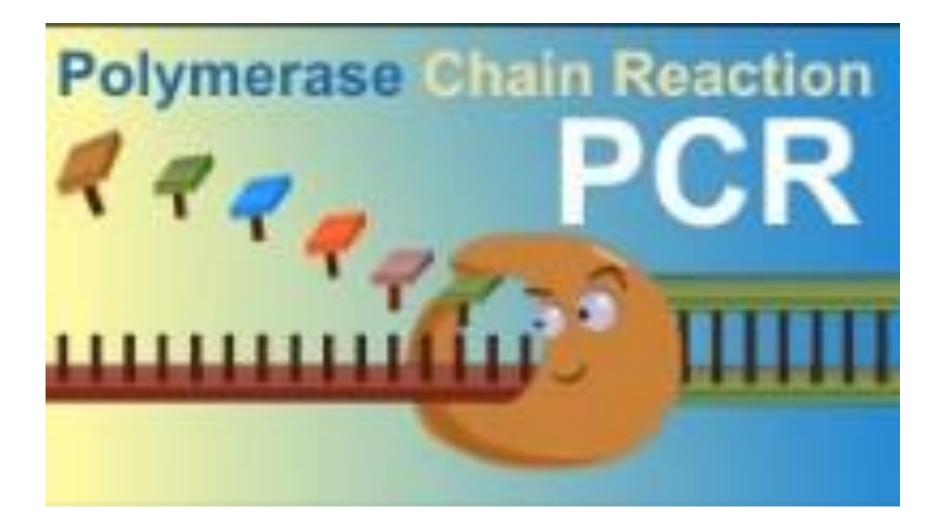
Polymerase chain reaction (PCR) based mapping

- Polymerase chain reaction (PCR) is a molecular biology technique and used to amplify specific regions of a DNA strand. This can be a single gene, just a part of a gene, or a noncoding sequence.
- It was Invented in 1983 by Dr. Kary Mullis. He later got Nobel prize for this invention

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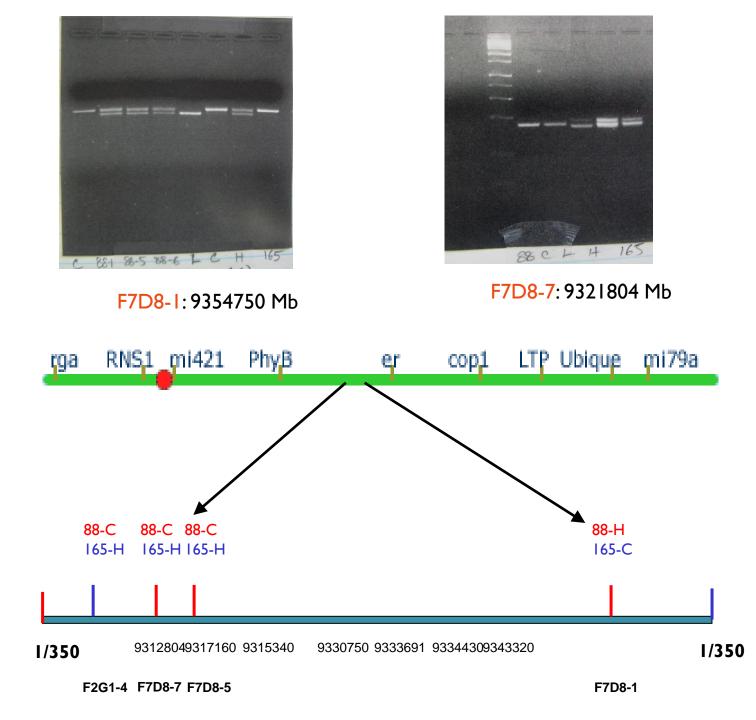




https://www.youtube.com/watch?v=iQsu3Kz9NYo







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1) Identify the mutation by sequencing

EIR1- chromosome 5 Marker- CTR1 and CIW14 Gene ID- At5g57090





2) Complementation test

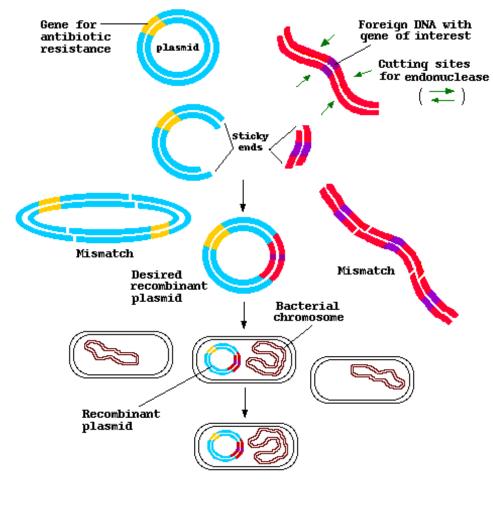
Inserting the wild-type gene in the mutant to see whether it can restore the abnormal phenotype





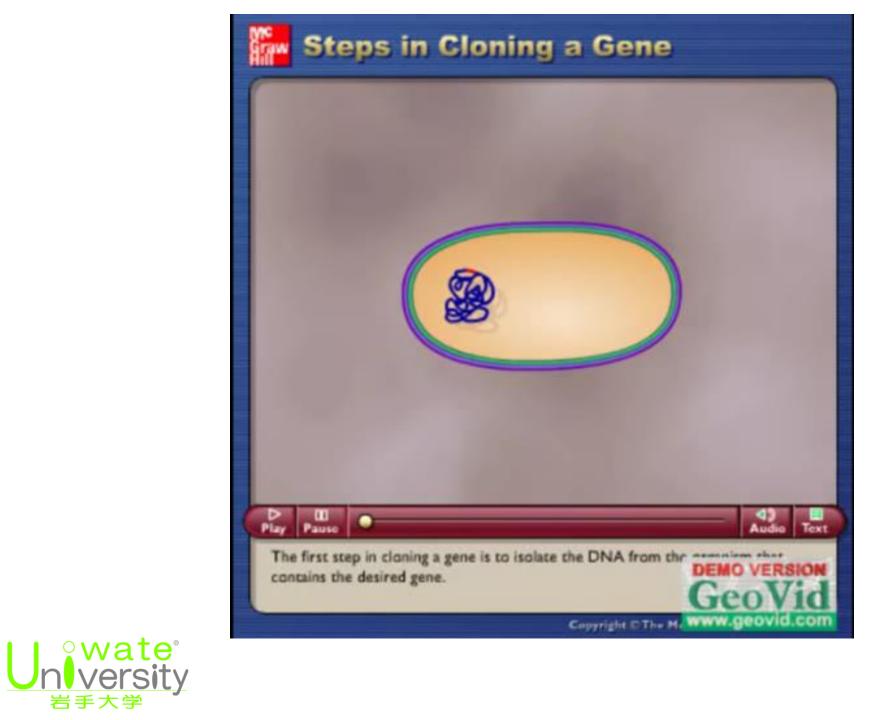
Steps in gene cloning

Plasmid Insertion









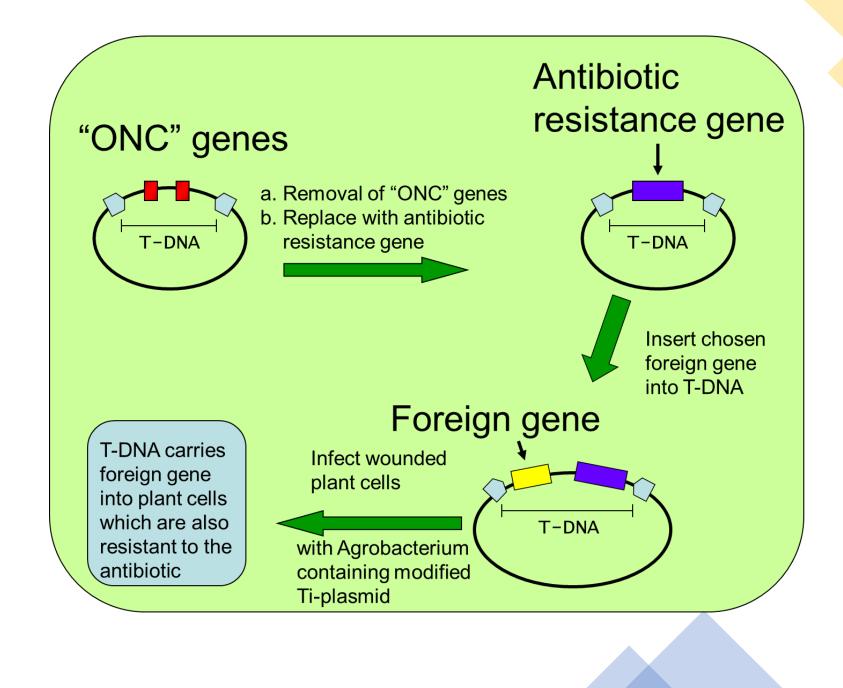


Once the gene is cloned in a vector, next step is to transfer the gene to the mutated plants

For transferring the gene, we use a natural soil bacteria, Agrobacterium and the process is called agrobacterium mediated transformation



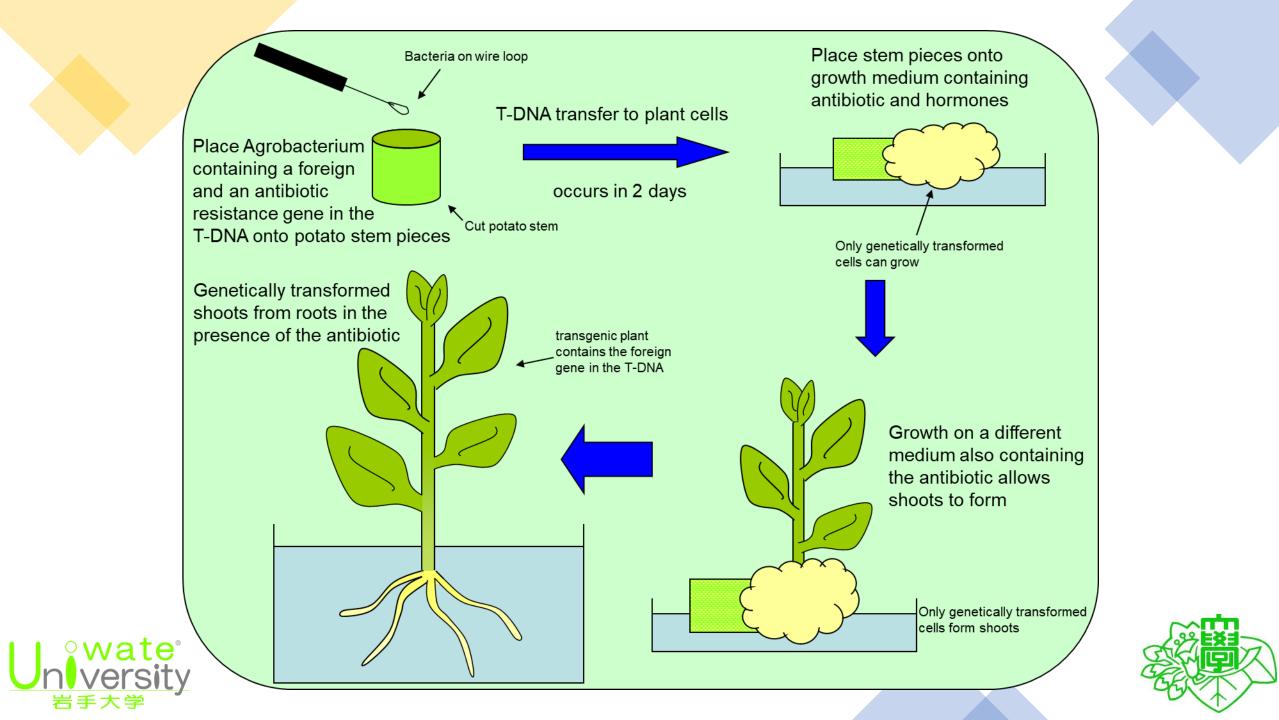




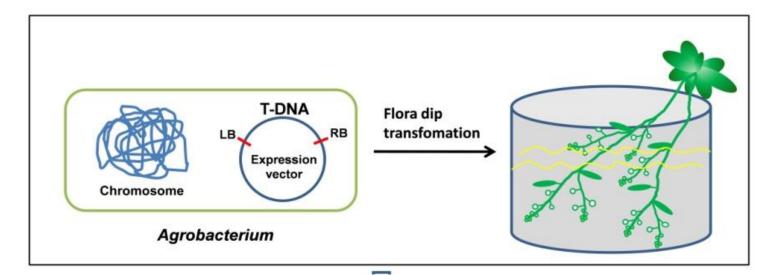
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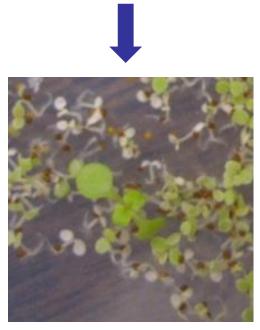
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Floral dip Transformation







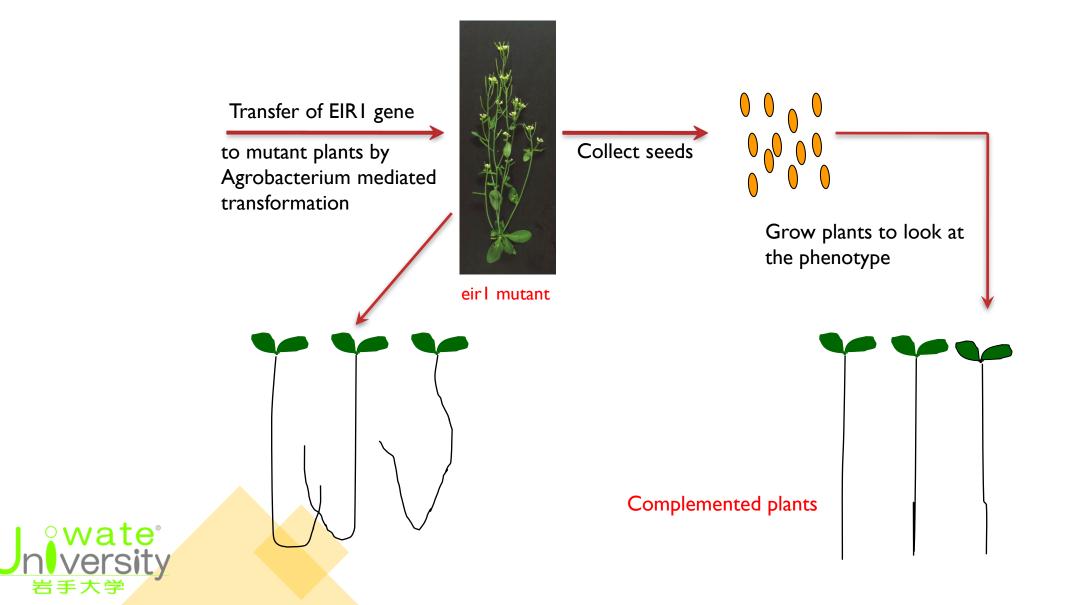
Grow plants to collect seeds



Screen with antibiotics



Final step: to validate the gene function, we need to restore the mutant phenotype





Real life application of gene transfer technique

By using this technique we can speed up our conventional breeding system

Enhance the nutritional value of the crops

Make the crops resistant to different adverse environmental conditions such as cold, drought, salinity, flood etc.

Bio-engineer the plants to detoxify the heavy metal contaminations such as cadmium, mercury, arsenic from soil

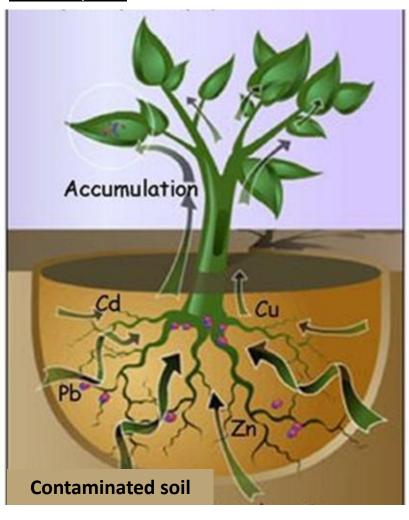
Create the exotic flowers with different colors and fragrance

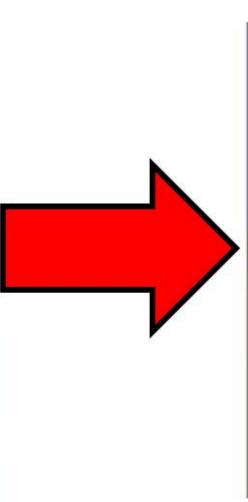
Produce vaccines in plant leaves



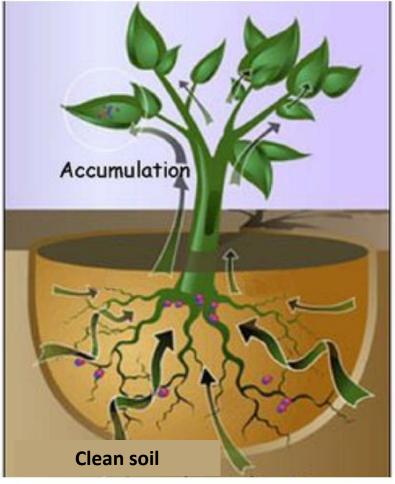
Concept of phytoremediation

Natural plant





Plants capable of efficiently transporting metals from soil

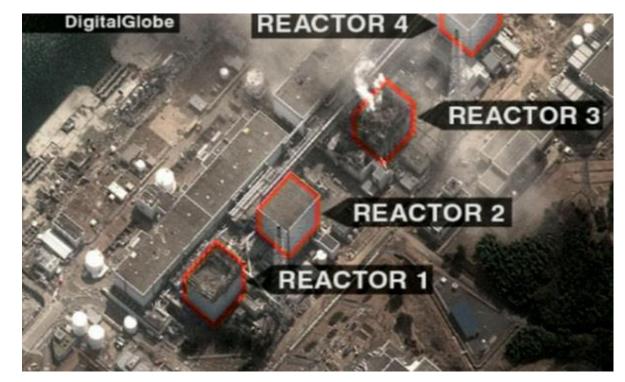








Chernobyl disaster (1986)



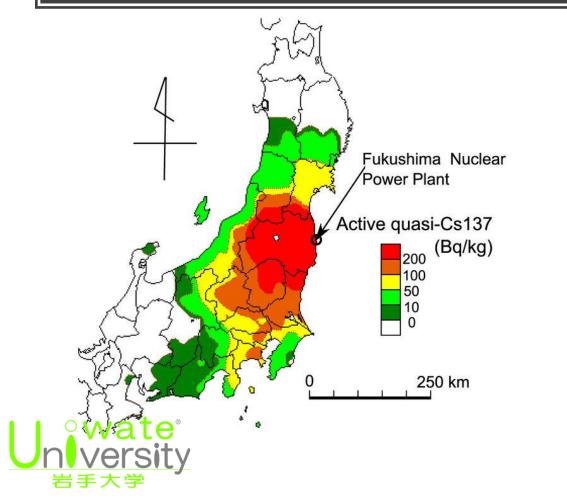
Fukushima disaster (2011)

Isotopes of Cesium $^{133}Cs \rightarrow Stable \text{ isotope}$ $^{134}Cs \rightarrow 2.0648 \text{ years half life}$ $^{137}Cs \rightarrow 30.17 \text{ years half life}$

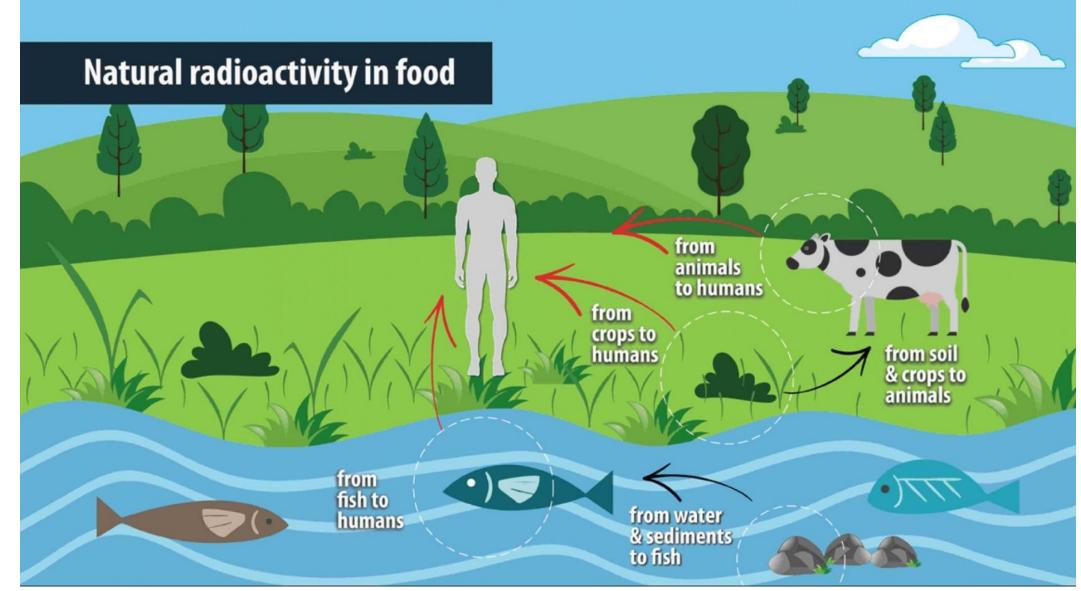




Radio cesium spreads from the nuclear plant area and finds its way towards river and agricultural land



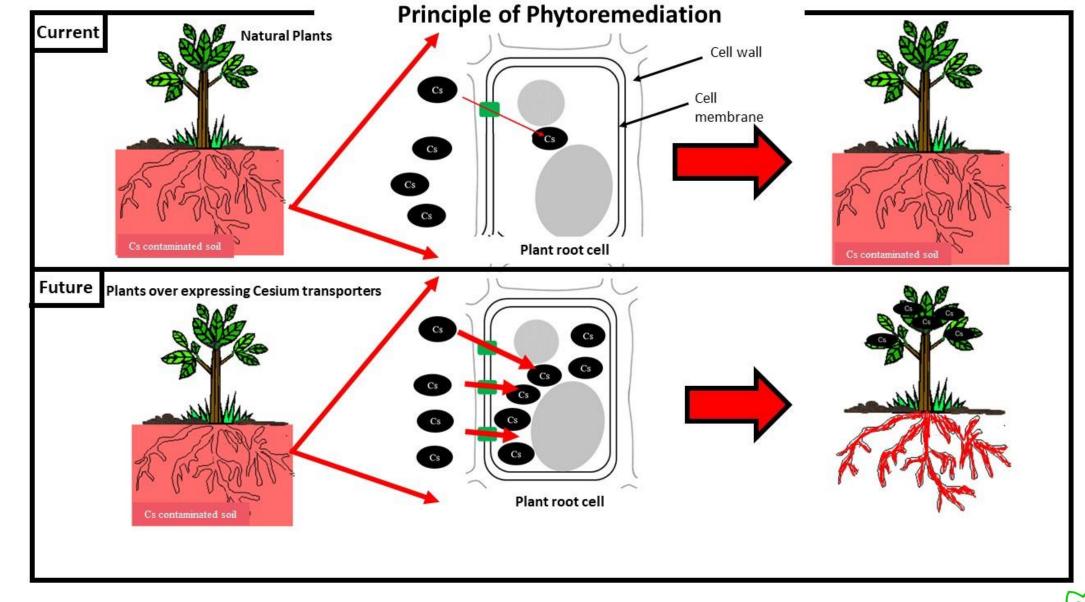
Chernobyl: Cs-137 Russ Belarus contamination 1090 square mile "exclusion zone" >104 Curies/sq mile onfiscated/Closed Zone eter than 40 curies per sous 5 to 15 Cifkm? of Cesam-1 3840 square miles strict radiation dose-control zone 39-104 Curies/sq mile



Radio cesium penetrates into our food chain

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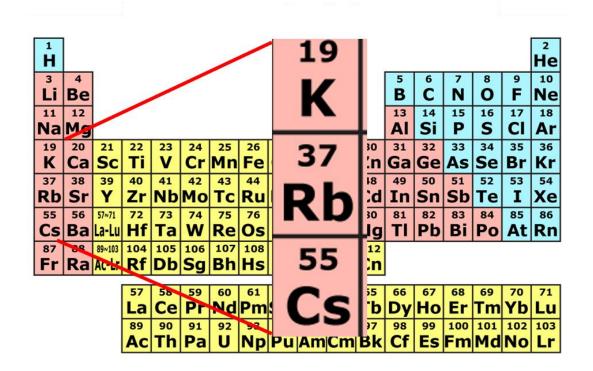


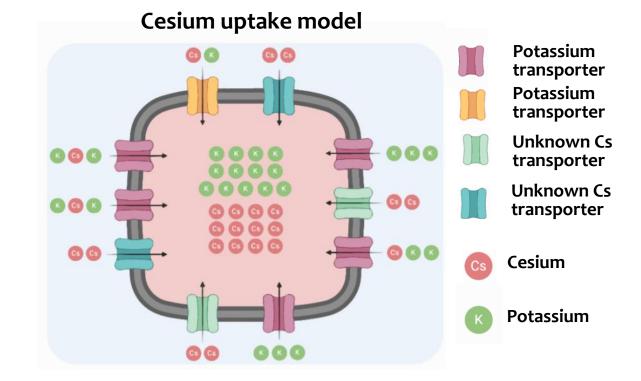
Can we clean up the radiocesium contaminated soil using plant?

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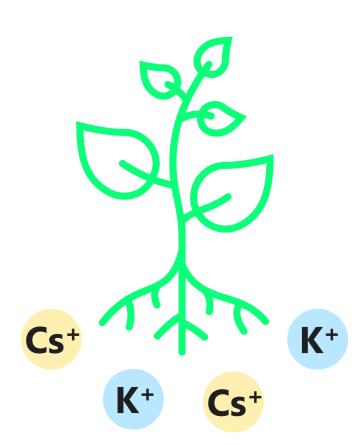






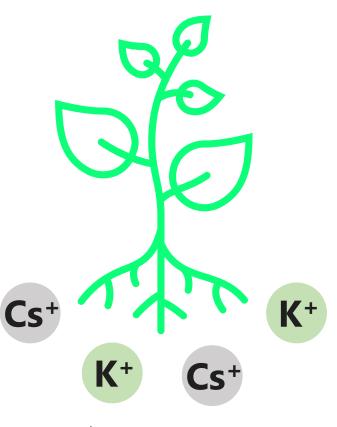


Cs⁺taken up by K⁺Transporters



K⁺-depleted soil



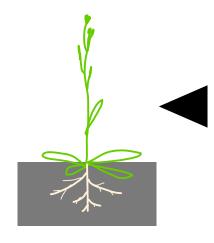


K⁺-independent

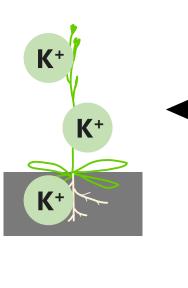
K⁺is retained in the soil



Are there K⁺-independent Cs⁺ transporters?

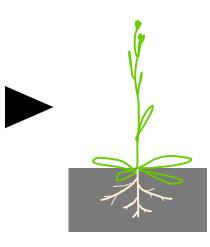


Several **ABC transporters** are differentially expressed



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The expression of these ABC transporters are not altered under K⁺

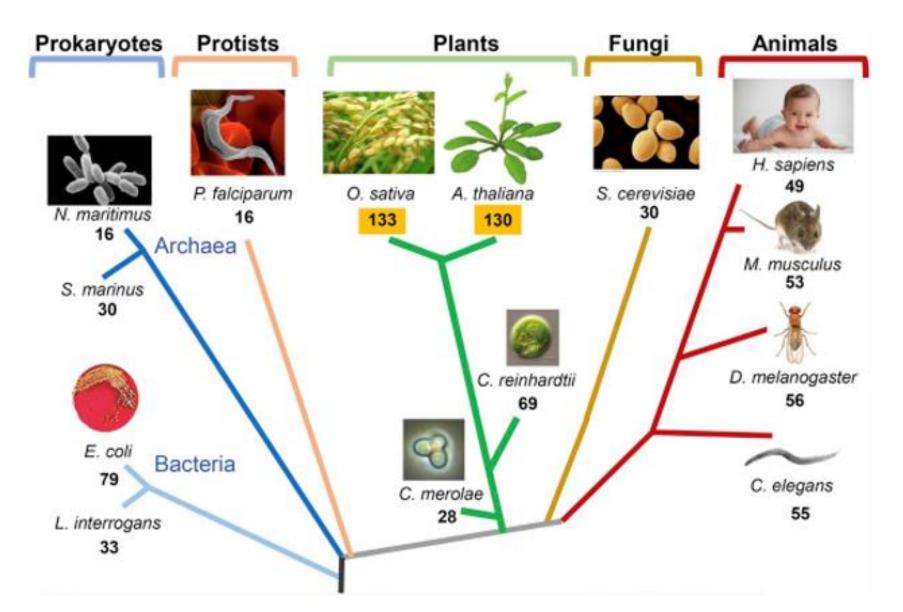


Cs⁺

Cs⁺



Hampton et al., Plant Physiology (2004) Microarray data





Reviewed Hwang et al., Molecular Plant (2016)

ABC transporters

ABCA	12
ABCB	30
ABCC	17
ABCD	2
ABCE	3
ABCF	5
ABCG	43
ABCI	16
Total	130

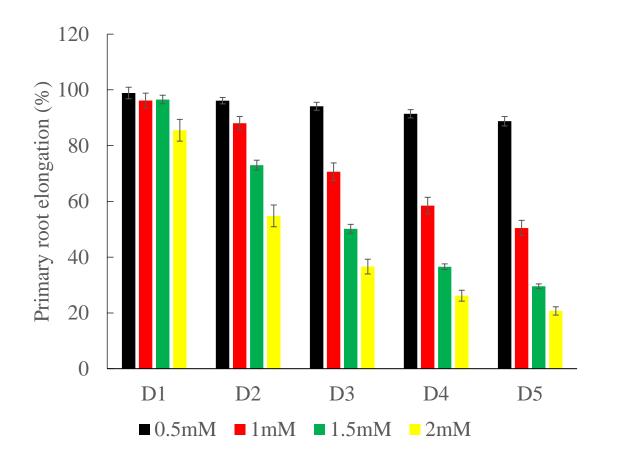
8 subfamilies

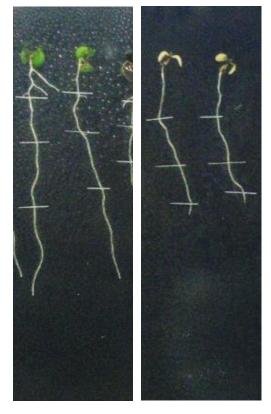
• Subfamilies **B**, **C**, and **G** \rightarrow <u>Metal transport activity</u>

 Numbers of each subfamily
of ABC transporters in Arabidopsis thaliana

Non-specific substrate selectivity of ABC transporters

Developing a root screening method



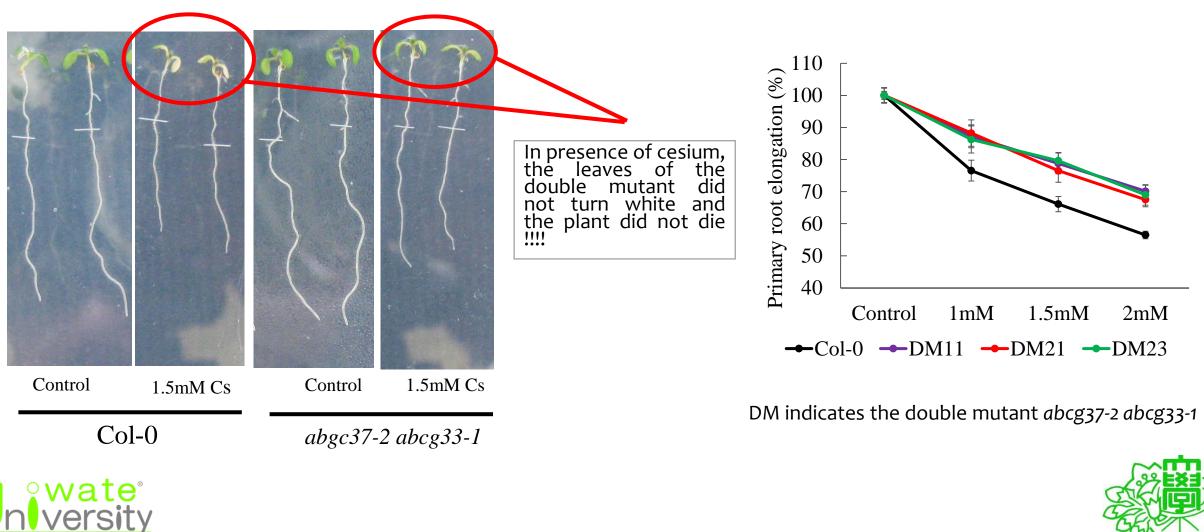


Control Cs



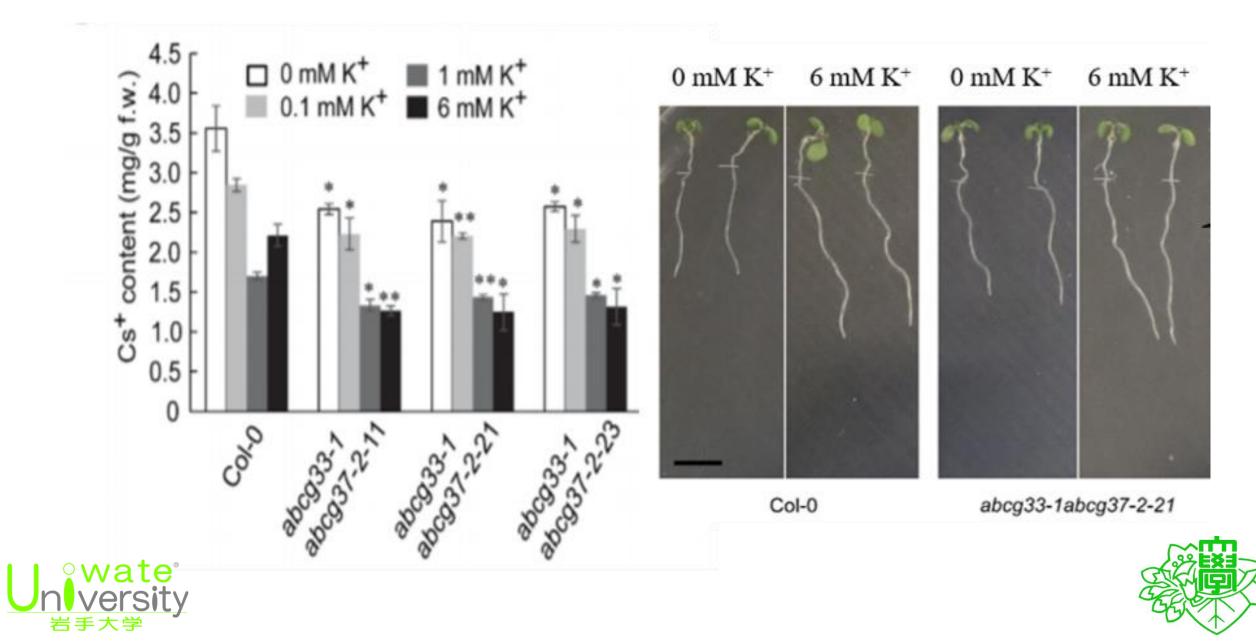


ABCG33 and ABCG37 possibly function as Cs⁺ transporters

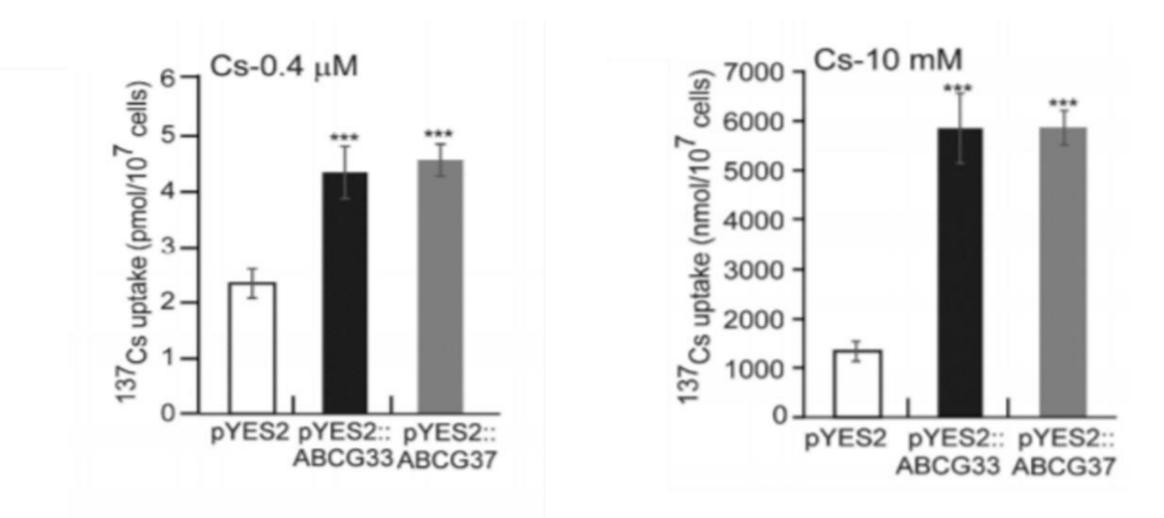




ABCG33 and ABCG37 are K⁺-independent Cs⁺ transporters



ABCG33 and ABCG37 are functional in yeast system and uptake Cs^+





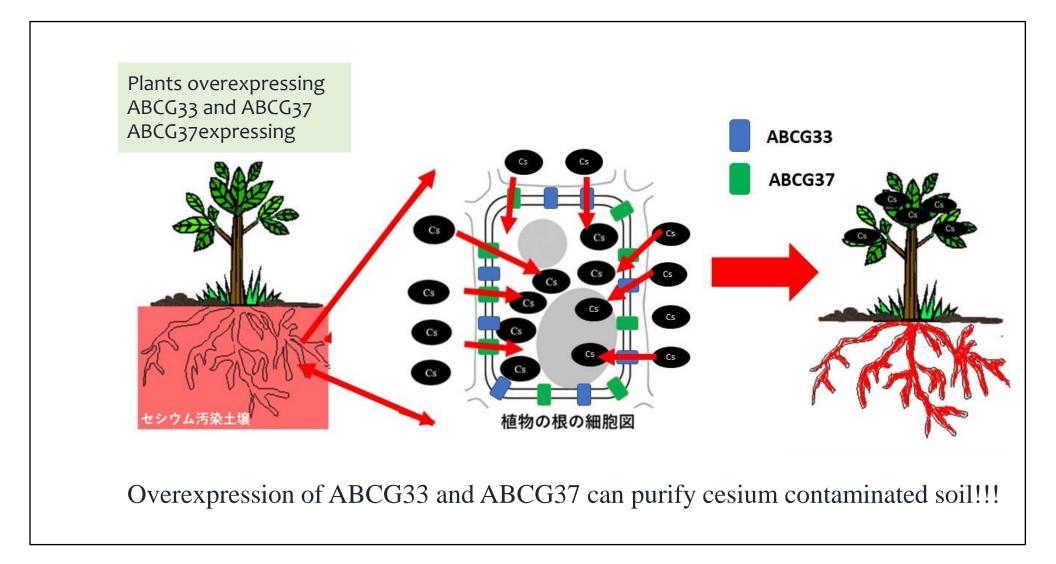


ABCG33 and ABCG37 do not uptake K^+

	1_	SD-gal BYT12 (trk1∆ trk2∆)				
	KCI (mM)	0	0.1	1.0	100	
	pYES2				0	Empty yeast vector
p	YES2::ABCG33					Yeast vector expressing ABCG33 and ABCG37
p	YES2::ABCG37	anter de la companya				
	pYES2::AtAKT1			0		Yeast vector expressing high affinity potassium transporter











Molecular Plant Q Register Claim Log in OsCNGC9 confer enhanced chilling non-canonical PAM compatibility in isolating genes tolerance in rice plants Wang et al. Li et al. Table of Contents > Wang et al. > View Archive -`ö́- About 🚼 Submit Alerts Se For Authors **Online Now** RESEARCH ARTICLE RESEARCH ARTICLE COMMENT RESEARCH ARTICLE

A transceptor-channel complex couples nitrate sensing to calcium signaling in *Arabidopsis* Wang et al. ATP Binding Cassette Proteins ABCG37 and ABCG33 function as potassium-independent cesium uptake carriers in Arabidopsis roots

Ashraf et al.

Breeding with Dominant Genic Male-Sterility Genes to Boost Crop Grain Yield in Post-Heterosis Utilization Era

Wan et al.

The landscape of gene-CDShaplotype diversity in rice (*Oryza sativa* L.): properties, population organization, footprints of domestication and breeding, and implications in





We got wide media coverage in Japan

International media:

AAAS's EurekAlert: tps://eurekalert.org/pub_releases/2021-02/iuj-pas021621.php

University press release:

Iwate University: <u>https://www.iwate-u.ac.jp/english/info/news/2021/02/003918.html</u> University of Tokyo: <u>https://www.a.u-tokyo.ac.jp/topics/topics_20210215-2.html</u>

Newspaper feature:

Microsoft News Japan: <u>http://a.msn.com/01/ja-jp/BB1dJ5Bv?ocid=sli</u>

FNN Prime Online: <u>https://www.fnn.jp/articles/-/145076</u>

Iwate Nippo: Attached image from the printed newspaper (<u>https://www.iwate-np.co.jp/article/2021/2/17/92217</u>) **TV channel coverage:** NHK: <u>https://www3.nhk.or.jp/lnews/morioka/20210216/6040009961.html</u>

Iwate Menkoi TV: <u>https://youtu.be/1q9z3hMmsKM</u>

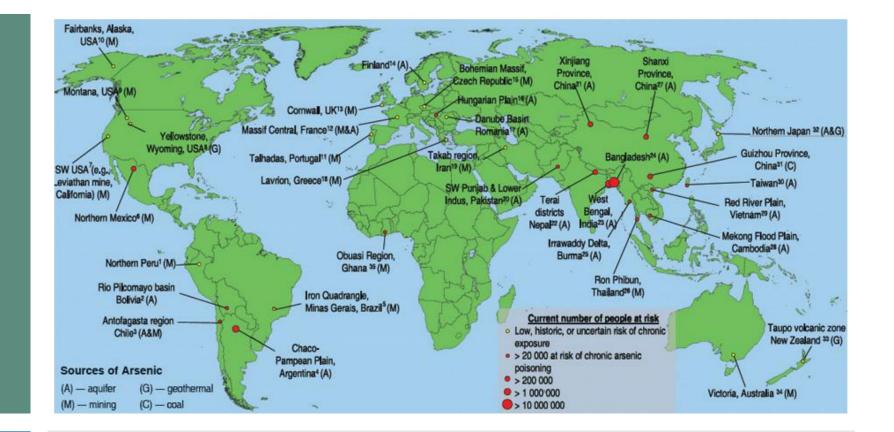




Global arsenic distribution

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- 140 million people in 50 countries have been drinking water containing arsenic at levels above the WHO provisional guideline value of 10 $\mu g/L$



•Arsenic is naturally present at high levels in the groundwater of a number of countries.

• Arsenic is highly toxic in its inorganic form.

•Contaminated water used for drinking, food preparation and irrigation of food crops poses the greatest threat to public health from arsenic.

•Long-term exposure to arsenic from drinking-water and food can cause cancer and skin lesions. It has also been associated with cardiovascular disease and diabetes.

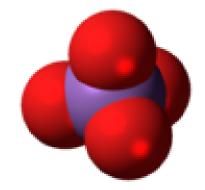




Inorganic Arsenic exists in two forms:

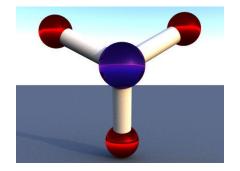
1) Fully oxidized pentavalent form: Arsenate

AsO₄³⁻ Phosphate analogue



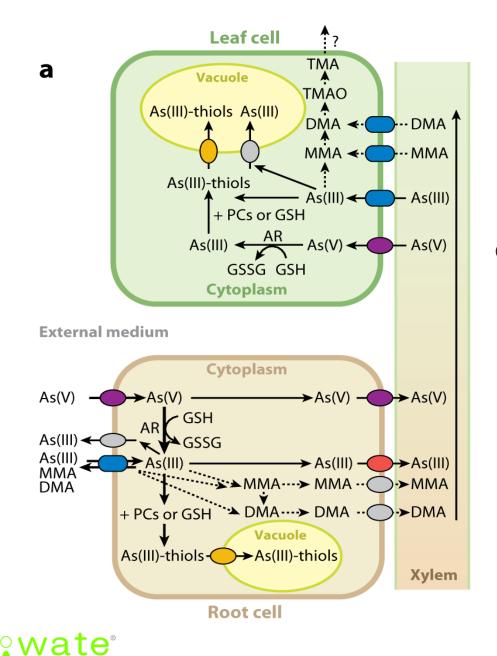
2) Reduced trivalent form: Arsenite

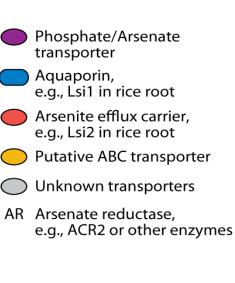
AsO₃³⁻











Arsenate is transported by phosphate transporters

Arsenite uptake is mediated by aquaporins

Arsenite efflux carrier is unknown



Only one protein LSi2 functions in cellular effflux of arsenite LSi2 is only present in rice, not in other higher plants

Possibility and Question:





What are the proteins???





EIR1, a highly conserved plant gene family with similarities to bacterial transporters

Particularly noteworthy is the similarity of EIR1 to the class of efflux carriers that remove toxic compounds from the interior of the cell. For example, *E. coll arsB* (P52146) represents a part of the arsenic efflux system (Diorio et al. 1995). *sbmA* (X54153), another integral membrane protein of *E. coll*, has been shown to be necessary for uptake of the antibiotic Microcin 25 (Salomon and Farias 1995). *Sortions* of EIR1 show 35%–40% similarity to these proteins. Our finding that the amino and the carboxyl terminus of EIR1 exhibit similarities to the

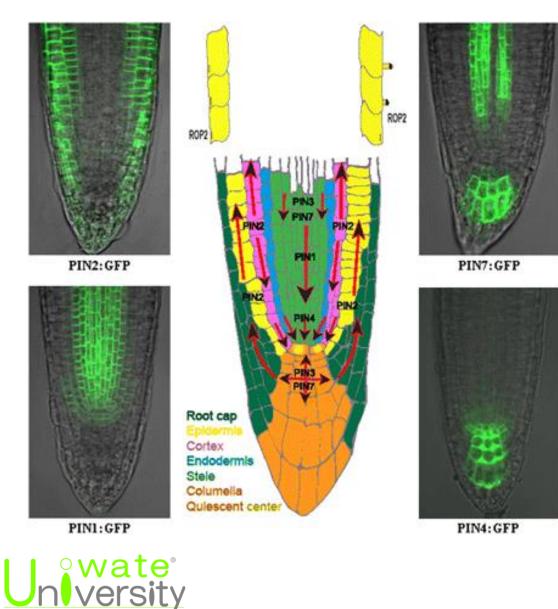
Luschnig et al., 1998; Genes &Dev

Portions of EIR1 show 35%-40% homology to bacterial arsenite transporter arsB

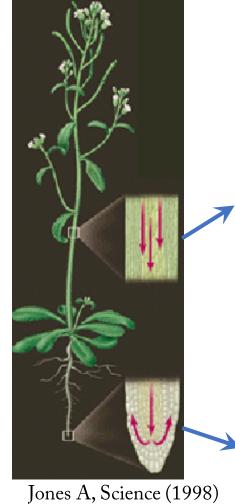




Auxin efflux carrier/PIN family proteins transport plant hormone auxin



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Acropetal transport (rootward transport)

 Basipetal transport (shootward transport)

Cladogram of plasma membrane residing PIN proteins







Identity matrix of Saccharomyces cerevisiae ACR3 (SsACR3), Pteris vittata ACR3 (PvACR3) and Escherichia coli arsenite transporter (arsB), Arabidopsis thaliana PINs (AtPIN1, AtPIN2, AtPIN3).

	SsACR3	PvACR3	arsB	AtPIN2	AtPIN1	AtPIN3
SsACR3	100.00	40.16	8.22	16.67	18.01	17.28
PvACR3	40.16	100.00	13.99	17.65	18.45	18.13
arsB	8.22	13.99	100.00	24.51	24.44	24.86
AtPIN2	16.67	17.65	24.51	100.00	62.88	60.88
AtPIN1	18.01	18.45	24.44	62.88	100.00	65.49
AtPIN3	17.28	18.13	24.86	60.88	65.49	100.00





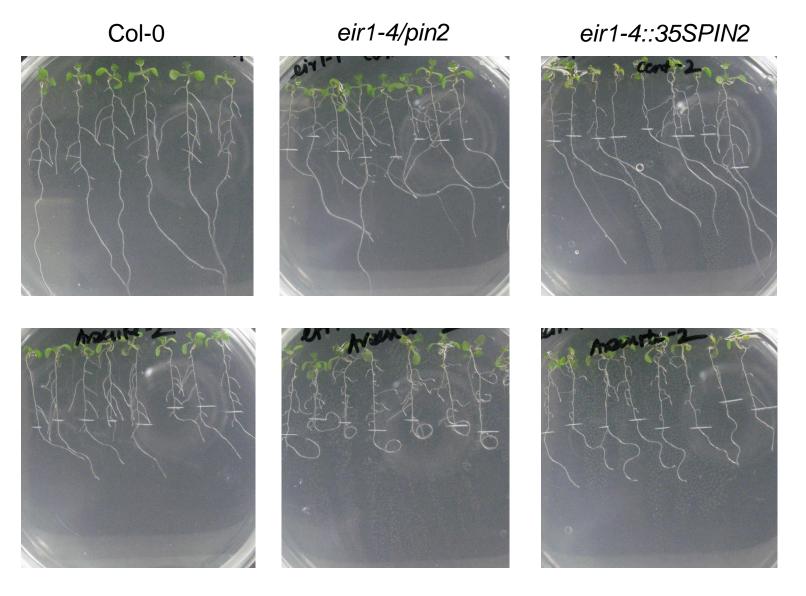
Identity matrix of OSLsi2 and arsenite transporters

	ScACR3	PvACR3	arsB	OsLsi2
ScACR3	100.00	40.87	10.89	9.27
PvACR3	40.87	100.00	10.19	7.43
arsB	10.89	10.19	100.00	14.29
OsLsi2	9.27	7.43	14.29	100.00





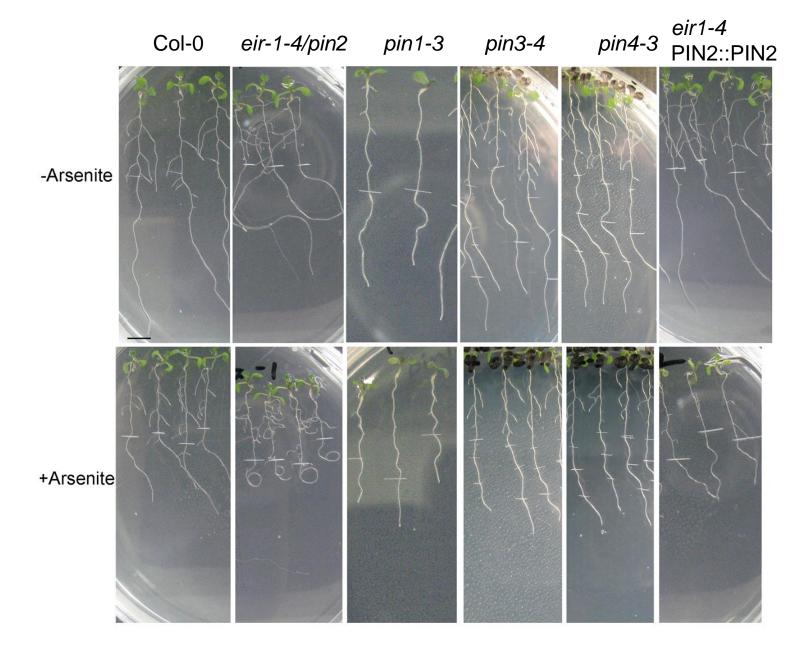
PIN2 mutant (pin2) shows hypersensitive response to arsenite







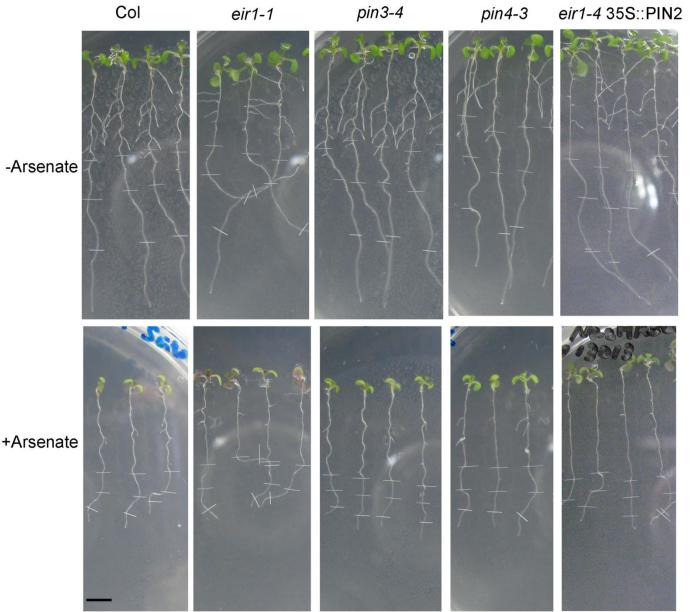
Other PIN mutants show wild-type like response to arsenite







pin2 mutant responds to arsenate like wild-type





+Arsenate



Summary



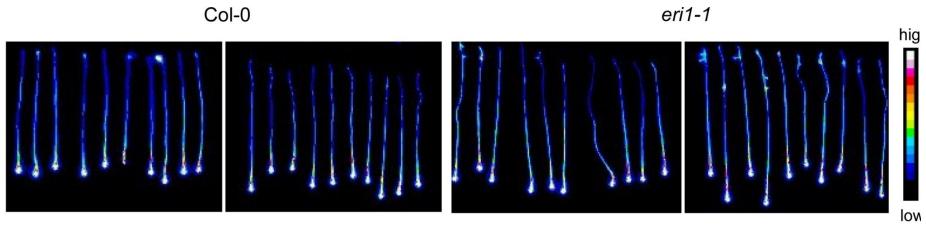
Auxin efflux carrier PIN2 regulates root arsenite but not arsentae response in Arabidopsis

Although all the PINs show similar homology to bacterial arsenite transporter arsB, only PIN2 is targeted by arsenite



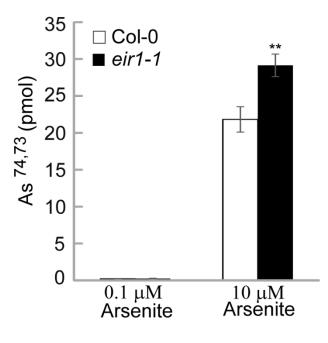


Arsenite transport is reduced in pin2 mutant



 $0.1 \ \mu M$ Arsenite

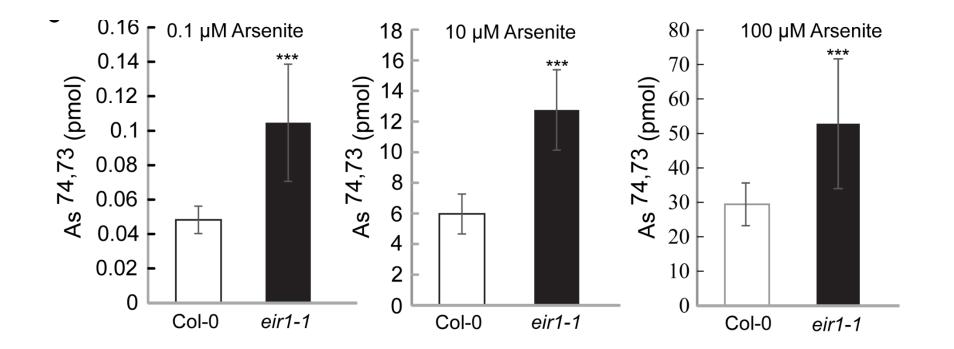








More Arsenite is accumulated in pin2 mutant







Elemental mapping by Synchotron Imaging



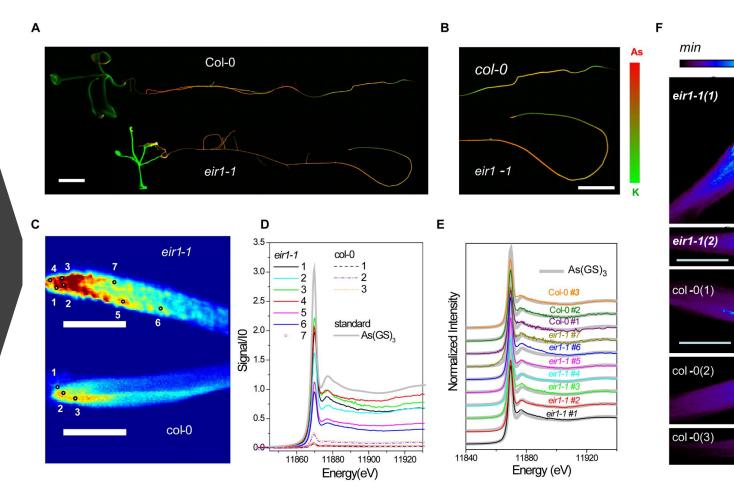




More Arsenite is accumulated in *pin2* mutant

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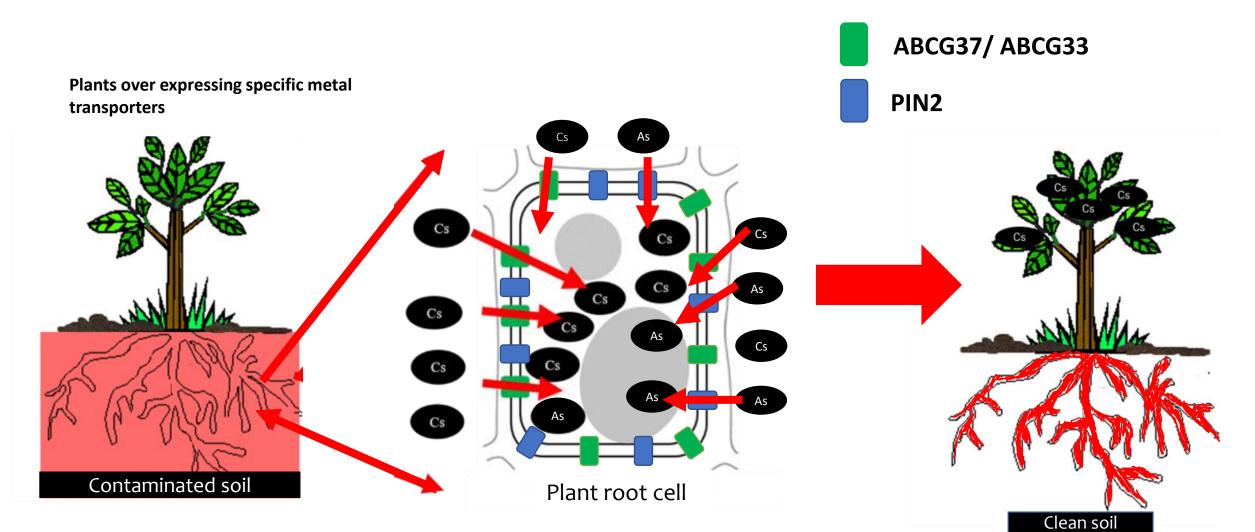
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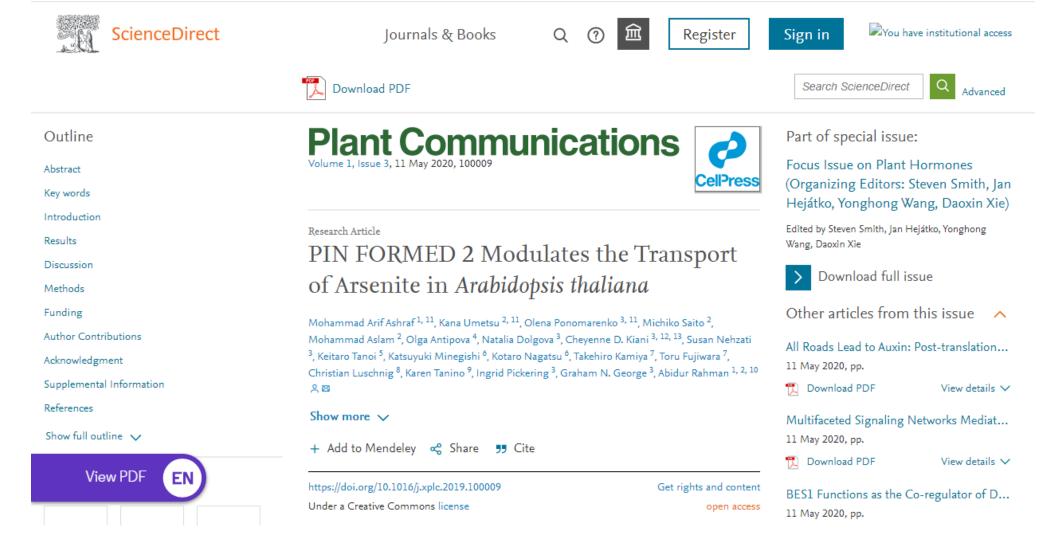
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Acknowledgements









Collaborators

Keitaro Tanoi Toru Fujiwara University of Tokyo

Graham George

Karen Tanino

University of

Saskatchewan















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